

Development of a Reverse Line Blot for simultaneous detection of Tick-Borne Pathogens in equines



V BLANDA, A CARUANA, S BONACCORSO, F LA RUSSA, R D'AGOSTINO, S VILLARI, G CAMILLERI, A TORINA, V FERRANTELLI

Istituto Zooprofilattico Sperimentale della Sicilia, Palermo, Italy

INTRODUCTION



Babesia caballi, *Theileria equi*, *Anaplasma phagocytophilum* and *Rickettsia* spp. are the main etiologic agents of Tick Borne Diseases (TBDs) in equines. Due to the nonspecific symptoms of TBDs, several molecular assays, each specific for a different Tick Borne Pathogen (TBP), are required for the diagnosis. This work aimed at developing a single Reverse Line Blot (RLB) for simultaneous detection and identification of equine TBPs.



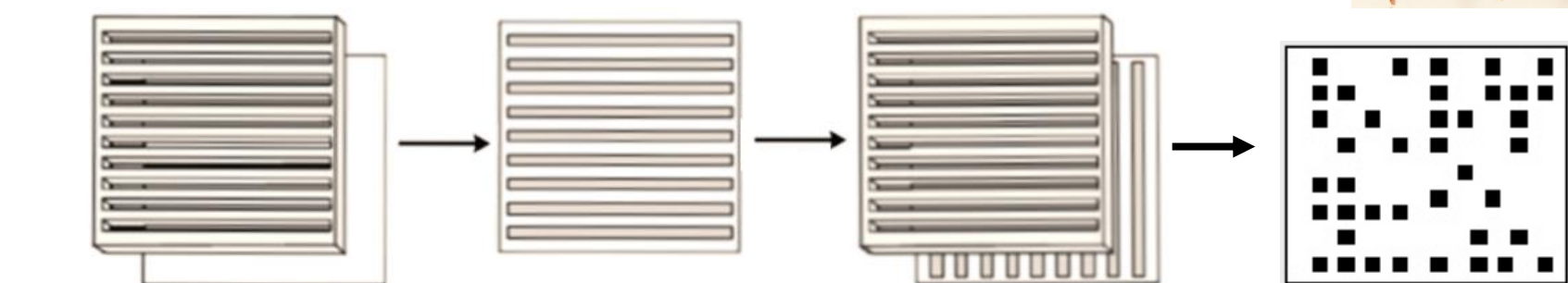
MATERIALS AND METHODS



PCRs with biotinylated primers on DNA from equine blood and ticks.

A new optimized multiplex PCR for *Anaplasma* spp. and *Rickettsia* spp. DNA amplification developed.

PCR products were diluted 1:4 in 2XSSPE / 0.1% SDS (a), to a final volume of 150 µl, heated for 10 min at 99°C (b), and immediately cooled on ice (c). Denatured PCR samples were placed in the slots (d). Excess of samples were aspirated (e). Hyperfilm preparation (f) and development in the darkroom was carried out (g).



Probe immobilization in a nitrocellulose membrane by a miniblotter.

Probes are bound covalently to the membrane

90° membrane rotation and application of biotinylated PCR products.

Hyperfilm exposure and development.



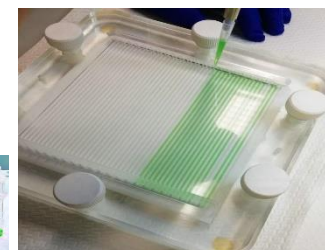
(a)



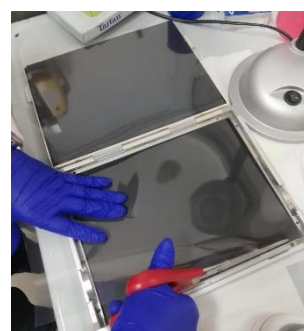
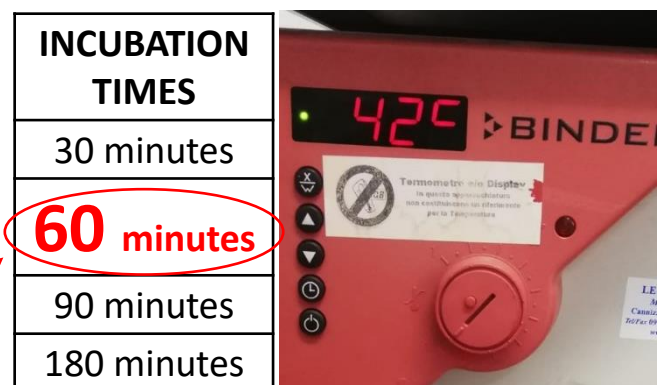
(b)



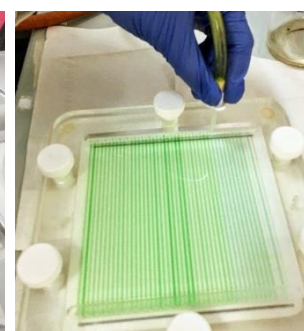
(c)



(d)



(e)



(f)



(g)

Sensitivity was determined by 10-fold serial dilutions of positive samples.

Specificity was tested with samples positive for other related pathogens.

Simultaneous detection of DNA from different pathogens was verified with different ratios in a single reaction mixture, loaded into a single channel.

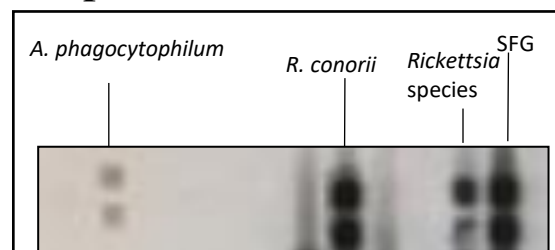
RESULTS AND CONCLUSIONS

The new RLB has been optimized with 15 different probes at two optimal concentrations:

PROBES	SEQUENCE	CONCENTRATION (pmol/150 µl)
<i>T. equi</i>	GTTTCGATTATTCTGTTCCCGG	1200;2400
<i>T. equi</i> -like	GGGGCATGTTTCATGACTCGA	1200;2400
<i>B. caballi</i>	GTTCGTTGTTCTTGGTTTTGCTT	1200;2400
<i>B. caballi</i> -like	CGGGTATTGACTTCGCTTTTTCTT	1200;2400
<i>Anaplasma</i> _HGE	GCTATAAAGAATAGTTAGTGG	1200;2400
<i>Ehrlichia</i> deer	GAATAATCTCTGAGCTGTCT	1200;2400
<i>R. sylvatica</i>	GTAGCCCTGCCACGATA	400;1200
<i>R. Conorii</i>	GTTATATACTGTAGCCCTG	400;1200
<i>R. aesch</i>	ATATTATACTGTATGAGCCCTG	400;1200
<i>R. helvetica</i>	CATGGCTTGATCCACGGTA	400;1200
Eq.pirolo_Catch all	TAATGGTTAATAGGARCRGTTG	1200; 2400
<i>A. phago</i> 1	TTGCTATAAAGAATAATTAGTGG	1200;2400
<i>A. phago</i> 2	TTGCTATRAAGAATARTTAGTGG	1200;2400
<i>Rickettsia</i> species	TAGCTCGATTGRTTACTTTG	400;1200
Spotted Fever Group <i>Rickettsia</i> (SFG)	ACTCACAARGTTATCAGGT	400;1200

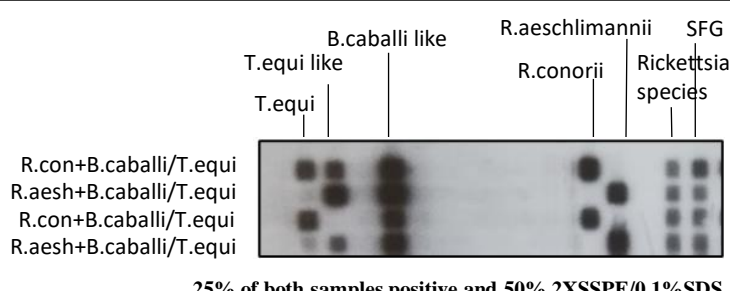
RLB sensitivity

Sensitivity assays produced a positive signal up to 1:1000 dilution for *Babesia*, *Theileria* and *Rickettsia* probes, and up to 1:10 for *Anaplasma* probes for both the probe concentrations.



The new multiplex PCR for *Anaplasma* and *Rickettsia* species made the test faster.

Amplicon combination in a single tube obtained by the different PCRs allowed the detection of different TBPs in a unique channel.

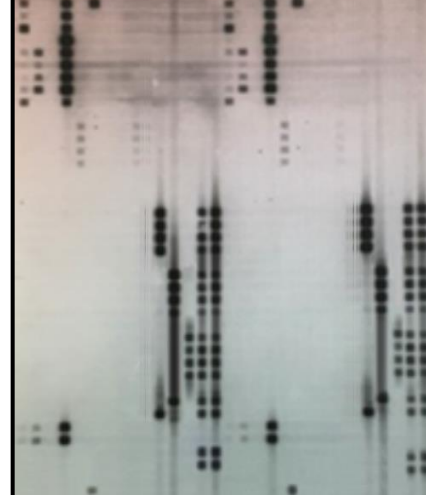


25% of both samples positive and 50% 2XSSPE/0.1%SDS

	1200 pmol/150µl					400 pmol/150µl					SFG			
	<i>T. equi</i>	<i>T. equi</i> like	<i>B. caballi</i>	<i>B. caballi</i> like	HGE	Catch all	<i>Ehrlichia</i>	<i>Anaplasma</i>	<i>Anaplasma</i> 2	<i>R. sylvatica</i>	<i>R. conorii</i>	<i>R. aesch</i>	<i>R. helvetica</i>	Generic
<i>T. equi</i> / <i>B. caballi</i>	•													
<i>T. equi</i> / <i>B. caballi</i>	•													
<i>T. equi</i> / <i>B. caballi</i>	•													
<i>T. equi</i> / <i>B. caballi</i>	•													
<i>T. equi</i> / <i>B. caballi</i> 1*	•	•												
<i>T. equi</i> / <i>B. caballi</i> 1:10	•	•												
<i>T. equi</i> / <i>B. caballi</i> 1:100	•	•												
<i>T. equi</i> / <i>B. caballi</i> 1:1000	•	•												
<i>T. equi</i> / <i>B. caballi</i> 1:10000	•	•												
<i>A. phagocytophilum</i>					•				•					
<i>A. phagocytophilum</i>					•				•					
<i>A. phagocytophilum</i> 1*					•				•					
<i>A. phagocytophilum</i> 1:10					•				•					
<i>A. phagocytophilum</i> 1:100					•				•					
<i>A. phagocytophilum</i> 1:1000					•				•					
<i>A. phagocytophilum</i> 1:10000					•				•					
<i>R. conorii</i> 1*										•	•	•	•	•
<i>R. conorii</i> 1:10										•	•	•	•	•
<i>R. conorii</i> 1:100										•	•	•	•	•
<i>R. conorii</i> 1:1000										•	•	•	•	•
<i>R. conorii</i> 1:10000										•	•	•	•	•
<i>R. aeschlimannii</i> 1*											•	•	•	•
<i>R. aeschlimannii</i> 1:10											•	•	•	•
<i>R. aeschlimannii</i> 1:100											•	•	•	•
<i>R. aeschlimannii</i> 1:1000											•	•	•	•
<i>R. aeschlimannii</i> 1:10000											•	•	•	•
<i>R. helvetica</i> 1*											•	•	•	•
<i>R. helvetica</i> 1:10											•	•	•	•
<i>R. helvetica</i> 1:100											•	•	•	•
<i>R. helvetica</i> 1:1000											•	•	•	•
<i>R. helvetica</i> 1:10000											•	•	•	•
<i>R. aeschlimannii</i>											•	•	•	•
<i>R. conorii</i>											•	•	•	•
<i>T. equi</i> / <i>B. caballi</i>	•	•												
<i>T. equi</i> / <i>B. caballi</i>	•	•												
<i>R. monacensis</i>														•
<i>R. felis</i>														•
<i>A. marginale</i>														•
<i>T. annulata</i>														•

First probes concentration

Second probes concentration



RLB Specificity

The probes used reacted only in correspondence of the DNA complementary to them demonstrating the specificity of the test.

*Pathogen DNA (10-30 ng)